

Executive editor: Vinod B. Shidham MD, FIAC, FRCPath Wayne State University School of Medicine, Detroit, MI, USA

Co-editors-in-chief:

Richard DeMay, MD (University of Chicago, Chicago, USA) Martha Pitman, MD (Harvard Medical School, Boston, USA) Vinod B. Shidham, MD, FIAC, FRCPath (WSU School of Medicine, Detroit, USA)

For entire Editorial Board visit : http://www.cytojournal.com/eb.pdf PDFs FREE for Members (visit http://www.cytojournal.com/CFMember.asp) **OPEN ACCESS** HTML format

p16^{INK4a} immunocytochemistry on cell blocks as an adjunct to cervical cytology: Potential reflex testing on specially prepared cell blocks from residual liquid-based cytology specimens

Vinod B. Shidham^{1*}, Ravi Mehrotra², George Varsegi³, Krista L. D'Amore³, Bryan Hunt¹, Raj Narayan⁴

Address: 1Department of Pathology, Wayne State University School of Medicine and Detroit Medical Center, Detroit, MI, USA, 2Department of Pathology, Moti Lal Nehru Medical College, University of Allahabad, Allahabad, India, ³Department of Pathology, Ex-cytopathology Fellow, Medical College of Wisconsin, Milwaukee, WI, USA, ⁴Department of Obstetrics and Gynecology, Medical College of Wisconsin, Milwaukee, WI, USA

E-mail: Vinod B. Shidham* - vshidham@med.wayne.edu; Ravi Mehrotra - rm8509@gmail.com; George Varsegi - varsegi@yahoo.com; Krista L. D'Amore -Krista.D'Amore@phci.org; Bryan Hunt: hunbry@yahoo.com, Raj Narayan: rnarayan@mcw.edu *Corresponding author

Published: 31 January 11

Cytolournal 2011, 8:1

Research Article

DOI: 10.4103/1742-6413.76379

Received: 27 June 10 Accepted: 10 January 11

This article is available from: http://www.cytojournal.com/content/8/1/1 © 2011 Shidham, et al.; licensee Cytopathology Foundation Inc.

This article may be cited as:

ShidhamVB, Mehrotra R,Varsegi G, D'Amore KL, Hunt B, Narayan R. p16 🕮 immunocytochemistry on cell blocks as an adjunct to cervical cytology: Potential reflex testing on specially prepared cell blocks from residual liquid-based cytology specimens. CytoJournal 2011;8:1 Available FREE in open access from: http://www.cytojournal.com/text.asp?2011/8/1/1/76379

Abstract

Background: p16^{INK4a} (p16) is a well-recognized surrogate molecular marker for human papilloma virus (HPV) related squamous dysplasia. Our hypothesis is that the invasive interventions and related morbidities could be avoided by objective stratification of positive cytologic interpretations by p16 immunostaining of cell block sections of cytology specimens. Materials and Methods: Nuclear immunoreactivity for p16 was evaluated in cell block sections in 133 adequate cases [20 negative for intraepithelial lesion or malignancy, 28 high-grade squamous intraepithelial lesion (HSIL), 50 low-grade squamous intraepithelial lesion (LSIL), 21 atypical squamous cells, cannot exclude HSIL (ASC-H), and 14 atypical squamous cells of undetermined significance (ASCUS)] and analyzed with cervical biopsy results. Results: (a) HSIL cytology (28): 21 (75%) were p16 positive (11 biopsies available — 92% were positive for cervical intraepithelial neoplasia (CIN) 1 and above) and 7 (25%) were p16 negative (3 biopsies available — all showed only HPV with small atypical parakeratotic cells). (b) LSIL cytology (50): 13 (26%) cases were p16 positive (12 biopsies available — all were CINI or above) and 37 (74%) were p16 negative (12 biopsies available — all negative for dysplasia. However, 9 (75%) of these biopsies showed HPV). (c) ASC-H cytology (21): 14 (67%) were p16 positive (6 biopsies available — 5 showed CIN 3/Carcinoma in situ/Ca and 1 showed CIN I with possibility of under-sampling. Cytomorphologic re-review favored HSIL) and 7 (33%) were p16 negative (5 biopsies available — 3 negative for dysplasia. Remaining 2 cases — 1 positive for CIN 3 and 1 showed CIN 1 with scant ASC-H cells on cytomorphologic re-review with possibility under-sampling in cytology specimen). (d) ASCUS cytology (14): All (100%) were p16 negative on cell block sections of cervical cytology specimen. HPV testing performed in last 6 months in 7 cases was positive in 3 (43%) cases. Conclusion: p16 immunostaining on cell block sections of cervical cytology specimens showed distinct correlation patterns with biopsy results. Reflex p16 immunostaining of cell blocks based on the algorithmic approach to be evaluated by a multiinstitutional comprehensive prospective study is proposed.

Key words: Cervical cancer, cervical cytology, immunohistochemistry, p16, Pap test, reflex testing, screening, squamous intraepithelial neoplasia

INTRODUCTION

Globally, after breast cancer, carcinoma of cervix is the second most frequent cancer in women.^[1] Detection of cervical premalignant lesions is a crucial component to reduce the associated morbidity and mortality. Over the years, cervical cytology (Pap test) has proven to be a very effective screening tool to achieve this goal, reducing the incidence from 14.8 per 100,000 in 1975 to 6.5 per 100,000 in 2006 in the United States^[2] and trends show a significant fall in incidence of 3.5% every year for the last ten years, until 2006.^[3] However, it is desirable to increase the specificity of this screening test. Any management algorithm that results in fewer false positives would be beneficial in preventing the complications of over-treatment.^[4]

p16^{INK4a} (p16), as a surrogate molecular marker of Human Papilloma Virus (HPV) related squamous dysplasia, has been relatively well established in cervical biopsy specimens^[5-8] with excellent inter- and intraobserver reproducibility.^[7] Its application to cervical cytology specimens has also been evaluated. However, the main challenge in our experience in applying p16 to cytology preparations is the inherent difficulty in interpretation of diagnostic nuclear immunoreactivity for p16 in whole cells with nucleus enveloped by surrounding cytoplasm due to the obscuring cytoplasmic immunoreactivity. As a result, reproducible and objective interpretation of p16 immunoreactivity in cytologic preparations especially with relatively few diagnostic intact cells is inherently suboptimal.^[9,10] Performing immunohistochemistry (IHC) on cell block sections of cytologic specimens could circumvent this limitation. However, preparation of cell blocks from liquid-based cytology (LBC) specimens with singly scattered cells may not produce cell block sections with reproducible cellularity.[11]

We applied a protocol specially standardized to produce cell blocks from cytology specimens with single scattered cells such as in LBC specimens.^[11] This protocol included a centrifugation step to align the dispersed solitary and small groups of cells in LBC specimens along the flat cutting surface of the cell block. A visible dark colored marker was included to monitor the depth of section cutting. By utilizing this protocol, the cell block sections showed a reproducible adequate cellularity. In this study, we evaluated application of p16 immunoreactivity in cell block sections prepared by this method from the residual Sure-Path[™] specimen (TriPath Imaging, Burlington, NC) (SP).

Our hypothesis is that ancillary reflex application of *p16 immunostaining on cell block sections of cytology specimens* could subcategorize *atypical squamous cells of undetermined significance* (ASCUS) and low-grade squamous intraepithelial lesion (LSIL) into cases with and without dysplasia. Similarly, atypical squamous cells, cannot exclude HSIL (ASC-H) and high-grade squamous intraepithelial lesion (HSIL) could be confirmed for dysplasia with increased objectivity. This approach would avoid invasive interventions and prevent potentially increased morbidities by utilizing objective options at the minimally invasive cervical cytology stage. Developing and adopting appropriate algorithms based on results of larger, comprehensive, multiinstitutional trial with reflex testing (p16 and other markers such as HPV L1 capsid protein^[12] and Ki67)^[13] on specially prepared cell blocks of cervical cytology specimens may be indicated.

MATERIALS AND METHODS

In this study, performed after approval from institutional review board (IRB), cell blocks were made from LBC specimens obtained from patients ranging 18 to 70 years of age. Out of approximately 120,000 gynecologic cytology specimens examined over a 2-year period, residual LBC specimens with positive cytopathologic interpretations (ASCUS and above) were available in 114 cases (in addition to 20 negative cases) for cell block preparation. The limiting factors controlling this final number included: (a) adequate cellularity of LBC, (b) availability of residual LBC specimen subject to reflex testing such as HPV test, (c) unavoidable logistic complexities related to selection of specimen for the study including multiplicity of cytopathologists-cytotechnologists involved with initial cytopathologic evaluation, (d) selection of representative numbers in each category of positive squamous lesions, and (e) practical limitations related to procurement of the residual specimen prior to its final disposal. These factors could not be controlled reproducibly to achieve inclusion of all cases based on limitations related to expedited IRB in smaller studies such as the present study.

Twenty cell blocks were prepared from cases with negative results and showing an uneventful follow-up pattern based on consecutive negative cervical cytologies and/or biopsies for one year. A total of 114 cell blocks were prepared from specimens with positive cytopathologic interpretations for squamous epithelial lesions (ASCUS and above). We used a standardized protocol for cell block preparation of specimens with singly scattered individual cells or small groups of cells using HistoGel[™].^[11] The protocol achieves alignment and concentration of the cells in the sample along the cutting surface of the cell block. It also includes AV-marker which serves to visualize the level at which the cells are concentrated. It allows selection of the sections from the plane of the cell block with the highest concentration of cells by the technologist during section cutting. For actual methodology, please refer to recently published video article which describes the technical details and is available as free publication in open access.[11]

Three unstained, serial, cell block sections (one

for hematoxylin and eosin (H & E) staining, one for p16 immunostaining, and one spare for elective immunostaining) were cut from each cell block. H & E stained sections were evaluated morphologically for cellularity. Cell blocks with sections showing more than 100 cells (either singly scattered or in cohesive groups) or showing any number of epithelial cells with morphological atypia were considered adequate. One (out of total 134) cell block was relatively hypocellular without morphological atypia and was not included in the study. The cytopathologic interpretations of 133 cases evaluated by p16 immunostaining of cell block sections included 20 negative for intraepithelial lesion or malignancy (NILM), 28 HSIL, 50 LSIL, 21 ASC-H, and 14 ASCUS [Figure 1]. One section from each of finally selected 133 cell blocks was immunostained for p16 [Table 1] with appropriate positive and negative controls for each batch.

http://www.cytojournal.com/content//8/1/1

The p16 immunoreactivity patterns included: (a) none, (b) cytoplasmic alone, (c) nuclear alone, and (d) nuclear with cytoplasmic. *Only nuclear and nuclear with cytoplasmic immunostaining were considered positive*. None or only cytoplasmic immunostaining was considered negative.^[10,14-16]

Cell block sections prepared from 20 cases with negative cytologic interpretation showed lack of nuclear (with or without cytoplasmic) immunoreactivity for p16 in squamous cells. Based on the previous study^[14] and many published studies,^[9,17-19] the p16 antibody clone used in this study (E6H4) showed excellent results.

The follow-up cervical biopsy results were available in 49 cases (15 out of 28 HSIL, 10 out of 21 ASC-H, and 24 out of 50 LSIL). Biopsy results were not available/performed in all 14 cases with ASCUS. *The cervical biopsy was considered*

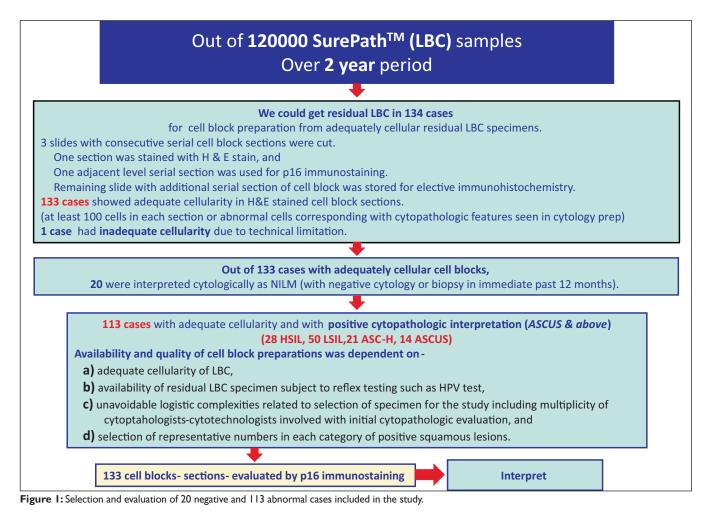


Table 1: Immunostaining of cell block sections with p16

| ······································ | | | | | |
|--|---------------------|------------------------------|----------|--|--|
| Clone | Source | Dilution | Duration | Pretreatment | |
| E6H4 | mtm laboratories AG | Proprietary (pre-diluted) | 30 min | Heat-induced epitope retrieval (35 min at 99°C, followed by 20 min cool off at room temperature) in citrate buffer, pH 6.0 | |

positive if it was reported as CIN1 or greater (confirmed with p16 IHC of biopsy sections in equivocal cases). Negative for dysplasia *with or without HPV cytopathic effects* were included for this study in the negative category.

The specimens were available for cell block preparation in 134 cases, because HPV DNA testing (HPVT) was not requested on the residual cervical cytology specimen. In cases with ASCUS interpretations, results of HPVT performed in last 6 months could be obtained in seven cases.

RESULTS

Correlation of p16 immunostaining on cell block sections of LBC specimens with biopsy results is shown in Table 2. The morphology of p16 immunostained cells in cell block sections in various groups is illustrated in Figures 2–4. Trouble shooting of false-positive and false-negative cases predominantly revealed sampling and biopsy interpretation issues. Cytoplasmic immunoreactivity alone was observed in scant cells in two cases.

I. Twenty-eight cases with HSIL cytology

- a. Twenty-one (75%) cases showed *positive results* with p16. Out of these, 12 had follow-up biopsies. Eleven (92%) biopsies were positive. The 75% (9/12) of these cases showed CIN 2–3 or invasive carcinoma (Ca), 2 showed CIN 1, and one was negative for dysplasia. Further analysis of three cases with CIN1 or lower lesions suggested potential undersampling in biopsy based on cyto–histo findings (with unequivocal cytomorphology for HSIL) and clinical details (such as with previous evidence of HSIL).
- b. Remaining seven (25%) HSIL cases showed *negative results* with p16. The cervical biopsy/cone results

were available in three cases, all of which showed only HPV changes without dysplasia (confirmed by p16 on biopsies). A review of initial cytology showed small atypical parakeratotic cells (SAPK)^[20] which were misinterpreted initially as HSIL cells (technically should have been ASC-H).^[20]

II. Fifty cases with LSIL cytology

- a. Thirteen (26%) cases showed *positive results* with p16. Out of these, 12 cases had biopsies, all of which were positive for CIN1 or above lesion.
- b. Remaining 37 (74%) LSIL cases showed *negative results* with p16. Cervical biopsies were available in 12 cases, all of which were negative for dysplasia but 9 (75%) of these biopsies showed the HPV cytopathic effect.

III. Twenty-one cases with ASC-H cytology

a. Fourteen (67%) cases showed *positive results* with p16.

Cyto-histo correlation was available during the period of study in six cases. Out of these, five cases showed CIN 3/carcinoma *in situ* (CIS)/Ca. The remaining one case showed CIN 1, but the cyto-histo correlation suggested that the lower grade findings in biopsy may have been related to sampling artifact with under-sampling. A review of cytology of all these ASC-H cases retrospectively showed cytomorphology favoring HSIL and could have been interpreted definitively as HSIL.

b. The remaining seven (33%) ASC-H cases showed *negative results* with p16. Cyto-histo correlation was available during the period of study in five cases. The 60% (3/5) were negative for dysplasia and 40% (2/5) were positive with CIN 3 in one and CIN 1 in the other. A review of initial cytology in both cases showed scant ASC-H cells and suggested the

| Cytology | p16 results on | Correlation with cervical biopsy | | | | |
|-------------------------|----------------|--|--|-------------------------|-------|--|
| | cell block | Positive cervical biopsy (CINI and above) | Negative cervical biopsy (HPV or negative for dysplasia) | Biopsy not available | Total | |
| HSIL (total cases 28) | Positive (21) | 11 | I | 9 | 21 | |
| | Negative (7) | 0 | 3 ª | 4 | 7 | |
| LSIL (total cases 50) | Positive (13) | 12 | 0 | I | 13 | |
| | Negative (37) | 0 | 12 | 22 | 37 | |
| ASC-H (total cases- 21) | Positive (14) | 5 | I | 8 | 14 | |
| | Negative (7) | 2 ^b | 3 | 2 | 7 | |
| ASCUS (total cases- 14) | Positive (0) | 0 | 0 | 0 | 0 | |
| | Negative (14) | 0 | 0 | 14 | 14 | |
| NILM (total cases- 20) | Positive (0) | 0 | 0 | 0 | 0 | |
| | Negative (20) | 0 | 0 | 20 | 20 | |

Table 2: Correlation of p16 immunostaining on cell block sections of LBC specimens with biopsy results

^aBiopsy showed HPV in all three cases with negative p16 on immunohistochemistry of biopsy. A reeview of initial cytology showed that the abnormal cells interpreted as HSIL were small atypical parakeratotic cells (SAPK).^[18]; ^bReview of initial cytology in both cases showed scant ASC-H cells and suggested sampling artifact in cell block sections immunostained for p16.

Figure 2: a) Pap smear interpreted as LSIL, b) H and E cell block sections, c) p16-stained cell block sections, d) biopsy showing CIN II-III.

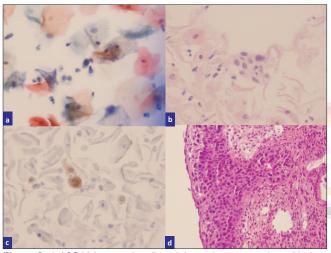


Figure 3: a) ASC-H (rare single cells with hyperchromatic nuclei and high N:C ratios), b) H and E stained cell block sections, c) p16-stained sections highlighting scattered high-grade cells, d) biopsy showing CIN III with extensive endocervical glandular involvement.

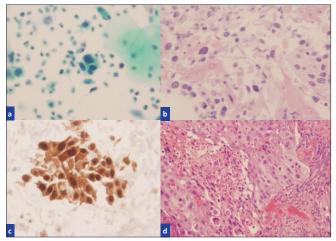


Figure 4: a) Pap smear interpreted HSIL, b) H and E cell block section containing "microbiopsies", c) p16-stained cell block section showing true nuclear positivity, d) biopsy showing invasive squamous cell carcinoma.

http://www.cytojournal.com/content//8/1/1

possibility of sampling artifact with the absence of representative abnormal cells in cell block sections immunostained for p16.

IV. Fourteen cases with ASCUS cytology

All 14 (100%) cases showed *negative results* and biopsies were not available in any. The total number of ASCUS cases could not be higher due to practical limitation of unavailability of samples in the majority of the cases, where the residual LBC was sent for reflex HPV testing. As the current specimens were not sent for HPVT during this ASCUS interpretation, they were available for cell block preparation for this study. Out of these 14 cases, results of HPVT performed in last 6 months were available in seven cases. Out of these seven cases, HPVT was positive in three (43%) cases.

Sensitivity and specificity were calculated only for definitive categories (HSIL and LSIL) using results on cases with unequivocal biopsy results [Tables 3 and 4]. The sensitivity for p16 in HSIL cases was 100% with specificity of 75%. The sensitivity for p16 in LSIL cases was 100% with specificity of 100% [Tables 3 and 4].

DISCUSSION

HPV is a proven carcinogen for cervical cancers and p16 is an excellent surrogate marker for HPV-related dysplasia.^[21] p16 is a cell-cycle inhibitor that binds to cyclin-dependent kinase 4 (CDK4) and prevents the phosphorylation and subsequent inactivation of the retinoblastoma protein (pRb).^[21] A reciprocal relation between p16 and pRb expression has been observed.^[22] Integration of high-risk HPV DNA into the host genome results in the overexpression of viral proteins E6 and E7. E7 binds to and inactivates pRB ultimately leading to overexpression of p16 through a negative feedback loop.^[23,24] The overexpression of p16 indicates already advanced interference of the viral oncoproteins with cellular proteins involved in cell cycle regulation. This phenomenon translates into nuclear immunoexpression of p16 in squamous epithelial cells and correlates with HPV-related dysplasia.[25,26]

A recent meta-analysis of 61 studies published until 2007 on p16 immunoexpression which included 27 studies on cytologic specimens and 34 studies on cervical biopsies.^[10] The analysis concluded that p16 immunostaining correlated with severity of cytological/histological abnormalities. However, the reproducibility was limited due to insufficiently standardized interpretation of the immunostaining. They recommended that a consensus needs to be reached for assessing p16 immunostaining. In addition, it needs to be assessed in various clinical settings addressing relevant management decisions.

Table 3: Sensitivity, specificity, positive predictive value, and negative predictive value for HSIL category with p16 immunostaining on cell block sections of 15 LBC specimens^a

| | Positive biopsy (CIN1 or above) | Negative biopsy (HPV alone or negative for dysplasia) | Total |
|--|---------------------------------|---|-------------|
| HSIL with p16 <i>positive</i> results on cell block (12) | II (TP) | I (FP) | 12 |
| HSIL with p16 negative results on cell block (3) | 0 (FN) | 3 (TN) | 3 |
| Total | 11 | 4 | 15 ª |
| Sensitivity TP/(TP+FN) | 1.00 | | |
| Specificity TN/(FP+TN) | 0.75 | | |
| PPV (positive predictive value) TP/(TP+FP) | 0.92* | | |
| NPV (negative predictive value) TN/(TN+FN) | I.00 [#] | | |

TP, True positive; FP, False positive; FN, False negative; TN, True negative; ^aOnly cases with cyto-histo correlation with biopsy were considered for these calculations; ^{*}The probability that the test would be True Positive is 0.9 and False Positive is 0.08; [#]The probability that the test would be True Negative is 1 and False Negative is 0

Table 4: Sensitivity, specificity, positive predictive value, and negative predictive value for the LSIL category with p16 immunostaining on cell block sections of 24 LBC specimens^a

| | Positive biopsy (CIN1 or above) | Negative biopsy (HPV alone or negative for dysplasia) | Total |
|---|---------------------------------|--|-------------|
| LSIL with p16 positive results on cell block (12) | 12 (TP) | 0 (FP) | 12 |
| LSIL with p16 negative results on cell block (12) | 0 (FN) | 12 (TN) | 12 |
| Total | 12 | 12 | 24 ª |
| Sensitivity TP/(TP+FN). | 1.00 | | |
| Specificity TN/(FP+TN). | 1.00 | | |
| PPV (positive predictive value) TP/(TP+FP) | 1.0* | | |
| NPV (negative predictive value) TN/(TN+FN) | I.00 [#] | | |

TP, True positive; FP, False positive; FN, False negative; TN, True negative; ³Only cases with cyto-histo correlation with biopsy were considered for these calculations;

*The probability that the test would be True Positive is I and False Positive is 0; #The probability that the test would be True Negative is I and False Negative is 0.

Out of 27 studies performed on cytological specimens, only 2 utilized cell block sections from residual cytology specimens.^[17,18] Remaining studies (n = 25) either utilized cytologic preparations or had not mentioned the details of preparation. Recently in 2010, one more study applied cell blocks to evaluate p16 immunoexpression in cytology specimens.^[27] Data from these studies also support our finding that p16 is a useful marker for the objective confirmation of CIN on cell blocks of cervical cytology specimens. However, cell block preparation methods were not standardized for achieving sections with good cellularity along the cutting surface of such cell blocks.^[11]

A review of these studies, surprisingly, highlights a critical flaw that the exact criteria for interpretation of p16 immunoreactivity as a marker of HPV-related dysplasias are significantly variable.^[10] Studies mostly support *nuclear immunoreactivity* with or without cytoplasmic immunostaining in morphologically squamous epithelial cells is consistent with HPV-related squamous dysplasia — irrespective of the number of cells.^[5-7,14,15,28]

Only 6 out of 61 studies in the meta-analysis spelled out the interpretation criteria for the p16 immunoreactivity pattern stressing nuclear immunoreactivity with or without cytoplasmic staining as diagnostic.^[10] Others, however, stated positive p16 immunoreactivity as nuclear and/or cytoplasmic staining implying that cytoplasmic immunoreactivity alone is also positive or else did not mention the criteria. This pattern of interpretation would obviously introduce potentially false-positive interpretations in such studies.

HPVT has been recommended as ancillary reflex test for managing ASCUS cytopathologic interpretations.^[29] Application of the HPVT in the concert with p16 will add further perspective as p16-positive and p16-negative cases supporting disruption or nondisruption of the cell cycle. If an HPVT is positive, then the p16-negative status indicates that the lesion is in a nondysplastic state, while the p16positive pattern indicates that the lesion is in a dysplastic phase. Such stratification may be of practical value especially in low risk postmenopausal women and adolescents.

Bose *et al.* reported that the highest rate of positivity (80%) and the highest levels of expression (more than three to five positive cells/10× field) were seen in HSIL and ASC-H cases. On the other hand, p16 positivity was noted in only 21% of LSIL and ASC-US cases. They concluded that,

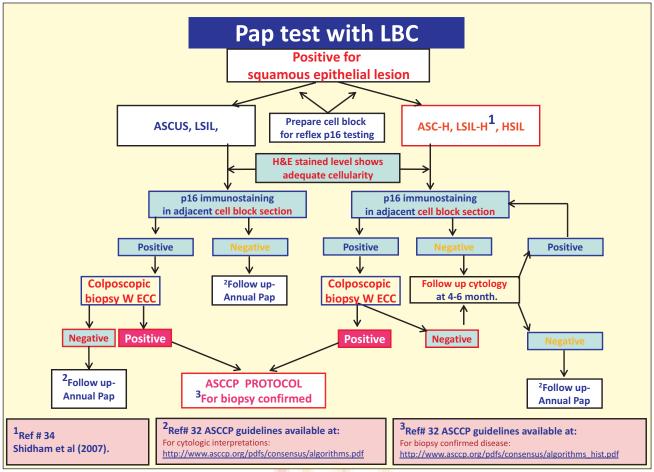


Figure 5: The recommended management algorithm using reflex p16 immunostaining on cell block sections of LBC specimens.

given that only a minority of LSIL cases progress on to higher-grade lesions, p16 might be useful for triaging these patients for closer follow-up and/or further evaluation.^[30] del Pino *et al.* in a recent report also reported comparative results on histopathology.^[31]

In our study, the majority of ASCUS (14 out of 14, 100%) and LSIL (37 out of 50, 74%) cases were negative for p16 in cytology specimens and in biopsy (equivalent to negative for dysplasia). Although this may appear as overdiagnosis, many of these cases were positive for either HPVT performed in the past 6 months (in 3 out of 7, 43% ASCUS cases) or the HPV cytopathic effect in biopsy (9 out of 12, 75% LSIL cases), consistent with HPV infection without dysplasia. If we consider this as overdiagnosis for dysplasia (not for HPV infection without dysplasia as the biological status) then ASCUS and LSIL cytopathologic interpretations although not uncommon, can be corrected with objective ancillary contribution by reflex p16 on cell block sections (Rp16) on the residual cervical cytology specimen. Based on the design of this study, concurrent HPVT on the same specimen could not be performed because the entire residual specimen was used for cell block preparation for p16 immunostaining.

The application of Rp16 on cell blocks of positive cytology specimens has very high potential of targeting selective intervention and preventing nonindicated invasive procedures by identifying p16 positive subset with increased probability of higher-grade lesions. This p16 assisted approach with reduced intervention should decrease the morbidity and cost associated with potential overtreatment.^[4] The p16 immunoexpression pattern in cytology specimens with positive cytologic interpretation for squamous epithelial lesions such as ASCUS, LSIL, ASC-H (including LSIL cannot rule out high grade (LSIL-H)^[32]), and HSIL could steer definitive management with significant savings and prevention of avoidable interventions. A comprehensive prospective ASCUS-LSIL Traige Study (ALTS) type multiinstitutional study evaluating Rp16 on cell block sections of residual cervical cytology specimens with positive results (ASCUS and above) in comparison to other alternatives including the HPV status would benefit patients and should be initiated with cost analysis. In addition to other benefits, such a prospective multiinstitutional study will achieve higher number of cases by including almost all potential specimens by preventing unavoidable exclusion of any of these relatively rare cases in routine setting similar to this study.

Currently, tests and procedures such as HPVT, colposcopy, and biopsy are applied to manage various cytologic interpretations as recommended by American Society for Colposcopy and Cervical Pathology (ASCCP).^[33] However, all these techniques are resource and manpower intensive, with related patient discomfort (colposcopy) and morbidity (cervical biopsy/conization) such as hemorrhage, cervical stenosis, cervical incomptence, and preterm delivery.^[4,34] HPVT is noninvasive, but it lacks specificity and only identifies the subset of cases with higher risk without any information-related to the absence or presence of dysplasia, which is a better decisive feature deciding the definitive management to minimize the invasive encounters. The specificity without decreasing the sensitivity of cervical cytology interpretations is highly desirable. This may be optimized by introducing the Rp16 as ancillary test^[35] on appropriately prepared cell block in all cases with positive cytologic interpretations. This allows an excellent noninvasive opportunity for appropriate decision making in the management algorithm. Rp16 on cell blocks of the residual LBC specimen in all positive cytology interpretations would achieve these features [Figure 5] and lead to significant savings with opportunity to decrease morbidity. The cell block protocol used in this study is an economical and easily available method.[11]

The cell block sections have additional benefits of sequential sectioning and immunostaining with other additional immunomarkers and future techniques including two color immunostaining.^[13,36] The location of AV marker in the section allows the application of the subtractive coordinate immunoreactivity pattern (SCIP) approach for proper evaluation of multiple immunomarkers in serial levels of cell block sections with singly scattered cells.^[37,38]

It is concluded that p16 on cell block sections of cervical cytology specimens with positive squamous interpretation including ASCUS, LSIL, ASC-H, LSIL-H, and HSIL showed distinct patterns with excellent correlation with biopsy results. Rp16 immunostaining of cell blocks prepared by a properly standardized protocol is recommended. An algorithmic approach with reference to p16 results to manage cytologically positive squamous lesions is proposed [Figure 5]. A multiinstitutional comprehensive prospective study to evaluate this approach is recommended as the next step to evaluate this algorithm with cost analysis.

List of abbreviations

ALTS, ASCUS-LSIL Triage Study; ASCCP, American Society for Colposcopy and Cervical Pathology; ASC-H, Atypical squamous cells Cannot exclude high-grade intraepithelial lesion; ASCUS, Atypical squamous cells of undetermined significance; Ca, invasive carcinoma; CDK4, cyclin-dependent kinase 4; CIN, cervical intraepithelial neoplasia; ECC, endocervical curettage; H & E, Hematoxylin and Eosin; HPV, human papilloma virus; HPVT, HPV DNA testing; HSIL, high-grade squamous intraepithelial lesion; p16, p16INK4a; IHC, immunohistochemistry; IRB, institutional review board; LBC, liquid based cytology; LSIL, lowgrade squamous intraepithelial lesion; LSIL-H, low-grade squamous intraepithelial lesion; cannot exclude HSIL; NILM, negative for *intraepithelial lesion or malignancy*; Pap test, cervical cytology; pRb, retinoblastoma protein; Rp16, Reflex p16; SAPK cells, small atypical parakeratotic cells; SCIP, subtractive coordinate immunoreactivity pattern; SP, SurePathTM specimen (TriPath Imaging, Burlington, NC).

ACKNOWLEDGEMENTS

This study was conducted at the Medical College of Wisconsin, (MCW) Milwaukee, WI, USA (during VS affiliation at MCW) and was presented in parts at different Annual Meetings of United States and Canadian Academy of Pathology, from 2004 through 2009. This article is a consolidation of these parts.

RM contributed to authorship during his participation in manuscript writing including a review of literature as an Indian Council of Medical Research (ICMR) Senior Visiting Faculty under mentorship of VB at the Department of Pathology, Medical College of Wisconsin, Milwaukee, WI, United States during March, 2010.

The authors appreciate and thank Glen Dawson, BS, HT, IHC(ASCP), Jerome Jacobson, HT, QIHC(ASCP), and Katherine Wertz, BS, HTL, QIHC(ASCP) for the technical immunocytochemistry assistance. We also thank Anushree Shidham for her secretarial and copy-editing support.

COMPETING INTERESTS

The author(s) declare that they do not have competing commercial interests.

AUTHORSHIP STATEMENT BY ALL AU-THORS

All authors of this article declare that we qualify for authorship as defined by ICMJE http://www.icmje.org/#author.

Each author has participated sufficiently in the work and take public responsibility for appropriate portions of the content of this article.

Each author acknowledges that this final version was read and approved.

ETHICS STATEMENT BY ALL AUTHORS

This study was conducted at Medical College of Wisconsin, Milwaukee, WI, USA with approval from Institutional Review Board (IRB) at Medical College of Wisconsin.

REFERENCES

- Jemal A, Siegel R, Ward E, Murray T, Xu J, Thun MJ. Cancer statistics, 2007. CA Cancer J Clin 2007;57:43-66.
- National Cancer Institute. Previous Version: SEER Cancer Statistics Review, 1975-2005 Available from: http://www.seer.cancer.gov/csr/1975_2006/ results_single/sect_05_table.04.pdf [Last accessed on 2010 Mar 14].
- Centers for Disease Control and Prevention. Cervical Cancer Trends. Available from:http://www.cdc.gov/cancer/cervical/statistics/trends.htm [Last accessed On 2010 Mar 14].
- Park SY, Bae DS, Nam JH, Park CT, Cho CH, Lee JM, et al. Quality of life and sexual problems in disease-free survivors of cervical cancer compared with the general population. Cancer 2007;110:2716-25,
- Chivukula M, Komorowski R, Shidham V. Ancillary application of p16^{INK4a} immunoexpression for objective grading of dysplasia in cervical biopsies. Mod Pathol 2004;17:1a-388.
- Chivukula M, Komorowski R, Shidham V. Application of p16^{INK4A} to evaluate the distribution pattern of mitotic figures and apoptotic bodies for grading cervical dysplasia. Mod Pathol 2004;17:1a-388.
- Behmaram B, Kotov P, Basir Z, Cafaro A, Novoa-Takara L, Shidham VB. Comparison of inter and intra-observer reproducibility of evaluating cervical dysplasia in HandE stained and p16 immunostained sections. Mod Pathol 2006;19:1a-359. Available from: http://www.nature.com/modpathol/journal/ v19/n1s/pdf/3800847a.pdf [Last accessed on 2010 Jun 12].
- Wentzensen N, Bergeron C, Cas F, Vinokurova S, von Knebel Doeberitz M. Evaluation of a nuclear score for p16INK4a-stained cervical squamous cells in liquid-based cytology samples. Cancer 2007;111:58-66,
- Wentzensen N, Bergeron C, Cas F, Eschenbach D, Vinokurova S, von Knebel Doeberitz M. Evaluation of a nuclear score for p16^{INK4a}-stained cervical squamous cells in liquid-based cytology samples. Cancer 2005;105:461-7.
- Tsoumpou I, Arbyn M, Kyrgiou M, Wentzensen N, Koliopoulos G, Martin-Hirsch P, et al. 2009 p16(INK4a) immunostaining in cytological and histological specimens from the uterine cervix: A systematic review and meta-analysis. Cancer Treat Rev 2009;35:210-20.
- Varsegi GM, Shidham V. Cell block preparation from cytology specimen with predominance of individually scattered cells. J Vis Exp 2009;29.doi: 10.3791/1316. PMID: 19623160 Video article is available FREE on web as open access from: http://www.jove.com/index/Details.stp?ID=1316. [Last accessed on 2010 Jun 12].
- Ungureanu C, Socolov D, Anton G, Mihailovici MS, Teleman S. Immunocytochemical expression of p16^{INK4a} and HPV L1 capsid proteins as predictive markers of the cervical lesions progression risk. Rom J Morphol Embryol 2010;51:497-503.
- Chivukula M, Austin RM, Duwe A, Friedman T, Masko J, Mauser N, et al. Dual-Stain for P16 and Ki67 in the Interpretation of Abnormal PAP Cytology Results: A Prospective Study. Mod Pathol 2010;23:85a-109. doi:10.1038/ modpathol.2010.10 Abstract #395. Available from: http://www.nature.com/ modpathol/journal/v23/n1s/pdf/modpathol201010a.pdf [Last accessed on 2010 Jun 12].
- Kotov P, Parameswaran L, Parisi JA, Chivkula M, Cafaro A, Fuentes M, et al. p16INK4a Immunostaing of Liquid Based Cervical Cytology Smears with SurePath®- Comparison of Two Antibodies. Mod Pathol 2005;18:1a-359. Available from: http://www.nature.com/modpathol/journal/v18/n1s/ pdf/3800908a.pdf [Last accessed on 2010 Jun 12].
- Kotov P, Parameswaran L, Parisi JA, Chivkula M, Cafaro A, Fuentes M, et al. Application of p16INK4A Immunostaing for Definitive Interpretation of ASC-H in Liquid Based Cervical Cytology Smears with SurePath ®. Mod Pathol 2005;18:1a-359. Available from: http://www.nature.com/modpathol/ journal/v18/n1s/pdf/3800908a.pdf [Last accessed on 2010 Jun 12],
- ShidhamV, D'Amore K, Varsegi G. Objective and definitive subcategorization of LSIL with p16INK Immunocytochemistry on Cell block Sections of Cervical Cytology Specimens. Cancer Cytopathol 2009;117:349-450.
- Liu H, Shi J, Wilkerson M, Huang Y, Meschter S, Dupree W, et al. Immunohistochemical detection of p16^{INK4a} in liquid-based cytology specimens on cell block sections. Cancer 2007;111:74-82.
- 18. Akpolat I, Smith DA, Ramzy I, Chirala M, Mody DR. The utility of $p \, I \, 6^{INK4a}$ and

http://www.cytojournal.com/content//8/1/1

Ki-67 staining on cell blocks prepared from residual thin-layer cervicovaginal material. Cancer 2004;102:142-9.

- Meyer JL, Hanlon DW, Andersen BT, Rasmussen OF, Bisgaard K. Evaluation of p16^{INK4a} expression in ThinPrep cervical specimens with the CINtec p16^{INK4a} assay. Cancer 2007;111:83-92.
- Chivukula M, Shidham V. ASC-H in Pap test- definitive categorization of cytomorphological spectrum. Cytojournal 2006;3:14. Free full text is Available from: http://www.cytojournal.com/content/3/1/14 PDF Available from; http:// www.cytojournal.com/content/pdf/1742-6413-3-14.pdf. [Last accessed on 2010 Jun 12].
- Kalof AN, Cooper K. p16^{INK4a} immunoexpression: Surrogate marker of highrisk HPV and high-grade cervica intraepithelial neoplasia. Adv Anat Pathol 2006;13:190-4.
- Tringler B, Gup CJ, Singh M, Groshong S, Shroyer AL, Heinz DE, *et al.* Evaluation of p16^{INK4a} and pRb expression in cervical squamous and glandular neoplasia. Hum Pathol 2004;35:689-96.
- Klaes R, Woerner SM, Ridder R, Wentzensen N, Duerst M, Schneider A, et al. Detection of high risk cervical intraepithelial neoplasia and cervical cancer byamplification of transcripts derived from integrated papillomavirus oncogenes. Cancer Res 1999;59:6132-6.
- Klaes R, Benner A, Friedrich T, Ridder R, Herrington S, Jenkins D, et al. p16^{INK4a} immunohistochemistry improves interobserver agreement in the diagnosis of cervical intraepithelial neoplasia. Am J Surg Pathol 2002;26:1389-99.
- Leversha MA, Fielding P, Watson S, Gosney JR, Field JK. Expression of p53, pRB, and p16 in lung tumours: A validation study on tissue microarrays. J Pathol 2003;200:610-9.
- Mäkitie AA, MacMillan C, Ho J, Shi W, Lee A, O'Sullivan B, et al. Loss of p16 expression has prognostic significance in human nasopharyngeal carcinoma. Clin Cancer Res 2003;9:2177-84.
- Yu L, Wang L, Zhong J, Chen S. Diagnostic value of p16INK4A, Ki-67, and human papillomavirus II capsid protein immunochemical staining on cell blocks from residual liquid-based gynecologic cytology specimens. Cancer Cytopathol 2010;118:47-55.
- 28. Varsegi G, D'Amore K, Shidham V. p16ink4a immunocytochemistry as an adjunct to cervical cytology potential reflex testing on specially prepared cellblocks from residual liquid based cytology (lbc) specimens. Mod Pathol 2009;22:p97a.Available from; http://www.nature.com/modpathol/journal/v22/n1s/pdf/modpathol2008212a.pdf [Last accessed on 2010 Jun 12].
- Stoler MH, Schiffman M. Interobserver Reproducibility of Cervical Cytologic and Histologic Interpretations: Realistic Estimates From the ASCUS-LSIL Triage Study. JAMA 2001;285:500-5.
- Bose S, Evans H, Lantzy L, Scharre K, Youssef E. p16(INK4A) is a surrogate biomarker for a subset of human papilloma virus-associated dysplasias of the uterine cervix as determined on the Pap smear. Diagn Cytopathol 2005;32:21-4.
- del Pino M, Garcia S, Fusté V, Alonso I, Fusté P, Torné A, et al. Value of p16(INK4a) as a marker of progression/regression in cervical intraepithelial neoplasia grade I.Am J Obstet Gynecol 2009;201:488.e1-7.
- 32. Shidham VB, Kumar N, Narayan R, Brotzman GL. Should LSIL with ASC-H (LSIL-H) in cervical smears be an independent category? A study on SurePathTM specimens with review of literature. Cytojournal 2007,4:7. Free full text is Available from: http://www.cytojournal.com/content/4/1/7 PDF Available from: http://www.cytojournal.com/content/pdf/1742-6413-4-7.pdf. [Last accessed on 2010 Jun 12].
- American Society for Colposcopy and Cervical Pathology guidelines Available from: http://www.asccp.org/pdfs/consensus/algorithms_cyto_07. pdf (ASCCP-Available from: http://www.asccp.org/hpv.shtml) [Last accessed on 2010 Mar 02].
- Luesley DM, McCrum A, Terry PB, Wade-Evans T, Nicholson HO, Mylotte MJ, et al. Complications of cone biopsy related to the dimensions of the cone and the influence of prior colposcopic assessment. Br J Obstet Gynaecol 1985;92:158-64.
- Wentzensen N, Bergeron C, Cas F, Vinokurova S, von Knebel Doeberitz M. Triage of women with ASCUS and LSIL cytology. Cancer 2006;111:58-66.
- Shidham VB, Varsegi G, D'Amore K. Two-color immunocytochemistry for evaluation of effusion fluids for metastatic adenocarcinoma. Cytojournal 2010;7:1.Available from:http://alturl.com/x4rg [Last accessed on 2010 Jun 12].

- Halloush RA, Akpolat I, Jim Zhai Q, Schwartz MR, Mody DR. Comparison of ProEx C with p16^{INK4a} and Ki-67 immunohistochemical staining of cell blocks prepared from residual liquid-based cervicovaginal material: A pilot study. Cancer 2008;114:474-80.
- ShidhamVB,Atkinson BF. Immunocytochemistry of effusion fluids: Introduction to the SCIP approach. In: Shidham VB, Atkinson BF, editors. 'Cytopathologic Diagnosis of Serous Fluids' Ch 5. 1st ed. Elsevier: W. B. Saunders Company; 2007. p. 55-78.

EDITORIAL / PEER-REVIEW STATEMENT

CytoJournal editorial team thanks the academic editor : Hormoz Ehya, M.D. Professor & Director of Cytopathology, Department of Pathology, Fox Chase Cancer Center, Philadelphia, PA, USA, for organizing and completing the double-blind peer-review process for this manuscript as per CytoJournal's peer-review policy. http://www.cytojournal.com/content/8/1/1

The FIRST Open Access cytopathology journal Publish in CytoJournal and RETAIN your *copyright* for your intellectual property Become Cytopathology Foundation Member to get all the benefits Annual membership fee is nominal US \$ 50 (US \$ 1000 for life) In case of economic hardship it is free For details visit http://www.cytojournal.com/CFMember.asp PubMed indexed FREE world wide open access

Online processing with rapid turnaround time. Real time dissemination of time-sensitive technology. Publishes as many colored high-resolution images Read it, cite it, bookmark it, use RSS feed, & many----







CYTOJOURNAL www.cytojournal.com



Peer-reviewed, Open access, Pub-Med indexed, Scholarly Cytopathology Journal

CytoJournal Editorial Board

CytoJournal International Editorial Panel



Barbara F. Atkinson, MD Exec Vice Chancellor, Kansas University Med Center, Kansas City, KS, USA
Editors-in-Chief
Executive-Editor

Editor-Emeritus



Richard M. DeMay, MD University of Chicago, Chicago, IL, USA



Martha B. Pitman, MD Harvard Medical School, Boston, MA USA



Vinod B. Shidham, MD, FRCPath, FIAC Wayne State Univ School of Medicine, Detroit, MI, USA CytoJournal Monographs

J Monograph Committee Co-chairs



Zubair Baloch, MD, PhD University of Pennsylvania Medical Center, Philadelphia, PA, USA



Shikha Bose, MD Cedars-Sinai Medical Center, Los Angeles, CA, USA CytoJ Monograph coeditors-in-chief



R. Marshal Austin, MD, PhD University of Pittsburgh Medical Center, Pittsburgh, PA, USA



Ruth Katz, MD M.D. Anderson Cancer Center, Houston, TX, USA



David C. Wilbur, MD Harvard Medical School, Boston, MA USA

Cytotechnology panel Jamie L. Covell BS, CT(ASCP) University of Virginia Health Sciences Center, Charlottesville, VA, USA Inderpreet Dhillon, MS, CT (ASCP) Detroit Medical Center, Detroit, MI, USA Gary W. Gill, Indianapolis, IN, USA Kalyani Naik, MS, SCT(ASCP) University of Michigan Hospital, Ann Arbor, MI, USA Consultant editors John N. Eble, MD, MBA

John N. Eble, MD, MBA Editorin-chief, Modem Pathology. Indiana University School of Medicine, Indianapolis, IN, USA. Stacey E. Mills, MD Editorin-chief, American Journal of Surgical Pathology University of Virginia Health Science Center, Charlottesville, VA, USA Mark R. Wick, MD Etitoric-thief, American Journal of Clinical Pathology.

Editor-In-chief, American Journal of Clinical Pathology University of Virginia Health Science Center, Charlottesville, VA, USA (Best in Cyto)/ Award Committee Chair Michael B. Cohen, MD Richard G. Lynch Chair of Experimental Pathology The University of Iowa, IA, US

Richard G. Lynch Chair of Experimental Pathology The University of Iowa, IA, US Founding Editor Vinod B. Shidham, MD, FRCPath, FIAC Wayne State University School of Medicine. Detroit Vinod B. Shidham, MD, FRCPath, FIAC

Associate Editors

Associate editors (Ad Hoc) Lester Layfield, MD ol of Medicine, Salt Lake City, UT, USA Eva M. Wojcik, MD Loyola University Medical Center, Chicago, IL, USA Associate editors (Rotating) Shikha Bose, MD ai Medical Center, Los Angeles, CA, USA David C. Chhieng, MD, MBA, MSHI Yale University, New Haven, CT, USA Mamatha Chivukula, MD niversity of Pittsburgh Medical Center, Pittsburgh, PA, USA Isam A. Eltoum, MD, MBA University of Alabama at Birmingham, Birmingham, AL, USA Rana S. Hoda, MD, FIAC New York, NY, USA dical Colle Nirag Jhala, MD, MIAC University of Pennsylvania Medical Center, Philadelphia, PA, USA Gladwyn Leiman, MBBCh FIAC FRCPath University of Vermont, Burlington, VT, USA Sanjay Logani, MD, MIAC NA LISA Sonya Naryshkin, MD, FIAC sville, WI, USA Liron Pantanowitz, MD University of Pittsburgh Medical Center, Pitts Husain Saleh, MD, FIAC, MBA Pittsburgh, PA, USA cine. Detroit. MI. USA Wayne Sate University School of Medicine, Momin T. Siddiqui, MD, FIAC Emory University, Atlanta, GA, USA Lourdes R. Ylagan, MD, FIAC Roswell Park Cancer Institute, Buffalo, NY, I NV USA

CytoJournal Editorial Board Members

R. Marshall Austin, MD, PhD (USA) Zubair Baloch, MD, PhD (USA) George Birdsong, MD (USA) Thomas A. Bonfiglio, MD (USA) Shikha Bose, MD (USA) David C. Chhieng, MD, MBA, MSHI (USA) Mamatha Chivukula, MD (USA) Douglas P Clark, MD (USA) Michael B. Cohen, MD (USA) Diane D Davey, MD (USA) Richard M DeMay, MD (USA) Hormoz Ehya, MD (USA) James England, MD, MBA (USA) Yener S Erozan, MD (USA) Prabodh Gupta, MBBS,MD, FIAC (USA) Amanda Herbert, MBBS, FRCPath (UK) Rana S. Hoda, MD (USA) Nirag Jhala, MD, MIAC (USA) Kusum Kapila, MD, FIAC, FRCPath (Kuwait) Ruth Katz, MD (USA) Sudha R Kini, MD (USA) Savitri Krishnamurthy, MD (USA) Leyster Layfield, MD (USA) Gladwyn Leiman, MBBCh FIAC FRCPath (USA) Virginia LiVolsi, MD (USA) Britt-Marie Ljung, MD (USA) Sanjay Logani, MD, FCAP, FASCP, MIAC (USA) Shahla Masood, MD (USA) Alexander Meisels, MD, FIAC (Canada) Dina R Mody, MD (USA) Sonya Naryshkin, MD, FIAC, FLAP (USA) Norimichi Nemoto, MD (Japan) Santo V Nicosia (USA) Svante R Orell, MD, FIAC (Australia) Martha B Pitman, MD ((USA) Liron Pantanowitz, MD (USA) David L Rimm, MD, PhD (USA) Husain Saleh, MD, FIAC, MBA (USA) Volker Schneider, MD, FIAC (Germany) Suzanne Selvaggi, MD (USA) Mark E Sherman, MD (USA) Vinod B Shidham, MD, FRCPath, FIAC (USA) Mary K Sidawy, MD (USA) Momin T. Siddiqui, MD, FIAC (USA) Jan F Silverman, MD (USA) Mark H Stoler, MD (USA) Rosemary Tambouret, MD (USA) Kusum Verma, MBBS, MD, MIAC (India) Philippe Vielh (France) David C Wilbur, MD (USA) Lourdes R. Ylagan, MD, FIAC (USA)

Argentina Ricardo Drut, MD (patologi@netverk.com.ar) Hospital De Ninos, La Plata, Argentina Boris Elsner, MD (betsner@elsitio.net) Argentina Lucrecia Illescas, MD, FIAC (illescas@fibertel.com.ar) Institub Papanicolaou, Buenos Aires, Argentina

Australia Andrew Field MB BS (Hons), FRCPA, FIAC, Dip of Cytopath (RCPA) (afield@stvincents.com.au) St Vincents Hospital, New South Wales, Australia

Belgium John-Paul Bogers, MD,PhD (John-Paul Bogers@ua.ac.be) University of Antwerp- Campus Groenenborger, Antwerp (Wirlik), Belgium Alain Verhest, M.D., PhD., FIAC (alain.verhest@bordet.be) Institut Jules Bordet, Brussels, Belgium Brazil

Joao Prolla, MD (jcprolla@yahoo.com) Hospital de Clinicas de Porto Alegre, RS, Brazil Vinicius Duval da Silva, MD, M.I.A.C. (vinids@pucrs.br / vinids@terra.com.br) Hospital Sao Lucas da PUCRS - IPB, Porto Alegre, Brazil Canada

Manon Auger, MD, FRCP(C) (manon.auger@mcgill.ca) McGill University Health Center, Montreal, PQ, Canada Diponkar Banerjee, MBChB,FRCPC,PhD (dbanerje@bccancer.bc.ca) BC Cancer Agency, Vancouver BC, Canada Terence J. Colgan, MD,FRCPC,FCAP,MIAC (tcolgan@mtsinai.on.ca) Mount Sinai Hospital, Toronto, ON, Canada

China Dongge Liu, MD/Phd (liudongge@sohu.com) Beijing Hospital, Beijing 100730, P. R. China

France Beatrix Cochand-Priollet, MD, Ph D, MIAC (beatrix.cochand-priollet@lrb.ap-hop-paris.fr) Lariboisière Hospital, Paris cedex, France Jerzy Klijanienko, MD (Jerzy.Klijanienko@curie.net) Institut Curie, Paris, France

Germany Magnus von Knebel Doeberitz, MD, PhD (knebel@med.uni-heidelberg.de) University of Heidelberg, Germany Ulrich Schenck MD (ulrich@schenck.de) Technical University of Munich, Munich, Germany

India Prakash Patil, MD, PhD (drprakash patil@yahoo.co.in) JM Medical College, Belgaum, India Arvind Rajwanshi, MD, MIAC, MNAMS, FRCPath (rajwanshiarvind@gmail.com) Post Graduate Institute of Medical Education & Research, Chandigarh, India Ravi Mehrotra, MD, MNAMS (rm850@ggmail.com) Moti Lal Nehru Medical College, Allahabad, India

Italy Guido Fadda, MD (Guidofadda@RM.unicattit) Catholic University - Largo Francesco Vito, Rome, Italy Pio Zeppa, M.D. (zeppa@unina.it) Università di Napoli "Federico II", Napoli, Italy

Japan Toshiaki, Kawai, MD (Ikawai@cc.ndmc.ac.jp) National Defense Medical College, Tokyo, Japan Robert Y, Osamura. MD

(osamura@isica.u-tokai.ac,ip) Tokai University School of Medicine, Kanagawa, Japan Jordan

Maher A. Sughayer, MD (msughayer@khcc.jo;msughair@hotmail.com) King Hussein Cancer Center, Amman, Jordan

Lebanon Nina Salem Shabb, MD (ns04@aub.edu.lb) American University of Beirut, Medical Center, Beirut, Lebanon

American University of Beirut, Medical Center, Beirut, Lebano Netherlands

Mathilde E. Boon, MD (m.e.boon@lcpl.nl) Leiden Cytology & Path Laboratory, Leiden, The Netherlands Pakistan Syed Mulazim Hussain Bukhari, MBBS, DCP, FCPS, Phd

(drmhbukhari@yahoo.com) King Edward Medical University, Lahore, Pakistan

Poland Wlodzimierz T. Olszewski, MD PhD (wtolszewski@coi.waw.pl) Institute of Oncology, Warsaw, Poland Portugal Margarida Almeida, MD (Margarida.Almeida@hsm.min-saude.pt) Hospital Santa Maria, Lisbon, Portugal

Evelina Mendonça, MD, MIAC (emendonca@polisboa.min-saude.pt) Insituto Portugués de Oncologia - Centro Regional de Lisboa, Lisboa, Portugal Fernando Carlos de Landér Schmitt, MD (Fernando.Schmitt@ipatimup.pt) da Universidade do Porto, Portugal

Singapore Alexander Russell Chang, MD (Otago), FRCPA, FHKCP (patarc@nus.edu.sg) National University of Singapore, Singapore Aileen Wee, MBBS, MRCPath, FRCPA (patwea@nus.edu.sg) National University Hospital, Singapore South Africa

South África Pam Michelow, MBBCh, MSc (Med Sci), MIAC (afainman@iafrica.com) National Health Laboratory Service (NHLS), Johannesburg, South Africa

Spain Jose M. Rivera Pomar, MD

(imivera@htru.osakidetz.net) Universidad Del Pais Vasco, Bilbao, Spain Mercedes Santamaria Martinez, MD (mersantamaria@medena.es) Hospital De Navarra, Pampiona, Spain Sweden

Annika Dejmek, MD, PhD (annika dejmek@pat.mas.lu.se) Malmö, Lund University, Malmo, Sweden Karin Lindholm, MD (karin.e. lindholm@telia.com) Malmo University Hospital, Malmo, Sweden Edneia tani (MD (edneia.tani@karolinska.se) Karolinska Hospital, Stockholm, Sweden Turkey

Binnur Uzmez Onal, MD, FEBP, FIAC (binnur@yahoo.com) SSK Training & Research Hospital, Ankara, Turkey Mehmet Akif Demir, MD (aki/demir@yaregion.se) Celal Bayar University, Manisa, Turkey

UK Minaxi S. Desai, MBBS, FRCPath (mina.desai@cmmc.nb.uk) Central Manchester & Manchester Children's University Hospital, Manchester, UK John H. F Smith, MD (John H. Smith@sth.nhs.uk) Royal Hallamshire Hospital, Sheffield, UK Allan Wilson, MD (Allan Wilson@lanarkshire.scot.nhs.uk) Manklands Hospital, Airdrie, UK

Uruguay Carmen Alvarez Santin, MD (alsanbla@adinet.com.uy) Laboratorio de Anatomia Patológica y Citologia. Facultad de Medicina. Montevideo, Uruguay

CytoJournal Virtual Trainee (CVT) Corner

(Under evolution) Lead section editor Walid E. Khalbuss, MD, PhD University of Pittsburgh Medical Center, Pittsburgh, PA, USA Section editors Zubair Baloch, MD, PhD University of Pernsylvania Medical Center, Philadelphia, PA, USA Guido Fadda, MD Catholic University - Largo Francesco Vito, Rome, Italy Liron Pantanowitz, MD University of Pittsburgh Medical Center, Pittsburgh, PA, USA

University of Pitssburgh Medical Center, Pittsburgh, PA, USA Paul Tranchida, MD Wayne State Univ School of Medicine & DMC, Detroit, MI, USA

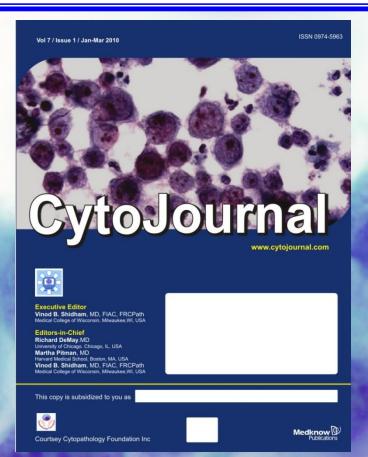
Medicolegal panel

Dennis R. McCoy, JD Hiscotk & Barday, Attorneys at Law, New York, NY, Mark S. Sidoti, Esq. Gibbons PC, New York, NY, Michael S. Berger, JD Andres & Berger, P.C., Haddonfield, NJ, USA Ken Gatter, MD, JD (gatterk@ohsu.edu) Oregon Health and Sciences University, Portland, OR

Statistical Advisor Varghese George, PhD (USA)

CytoJ OA Advocacy Committee Chair Lynn Sandweiss, MPH (lynn.sandweiss@gmail.com)

Managing Editor Anjani Shidham, BS (USA)



Open Access cytopathology journal

Publish in CytoJournal and RETAIN your copyright for your intellectual property

Become Cytopathology Foundation Member to get all the benefits Annual membership fee is nominal US \$ 50 (US \$ 1000 for life) In case of economic hardship it is free For details visit http://www.cytojournal.com/CFMember.asp

PubMed indexedFREE world wide open accessDouble blind peer review processOnline processing with rapid turnaround time.Real time dissemination of time-sensitive technology.Publishes as many colored high-resolution imagesRead it, cite it, bookmark it, use RSS feed, & many----



CYTOJOURNAL

www.cytojournal.com Peer-reviewed academic cytopathology journal

